

Tag SNP selection in genetic association studies

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Power & efficiency of association studies

- Statistical power of association studies increases with the number of individuals and the density of SNPs being genotyped.
- Genotyping cost (efficiency) is affected by the overall number of genotyped SNPs.
- Select a minimal subset of markers (**tag SNPs**) that predict remaining SNPs (**target SNPs**) with high accuracy.

“Predict a SNP”

Hap1	A	G	T	A
Hap2	A	C	A	C
SNP #	1	2	3	4

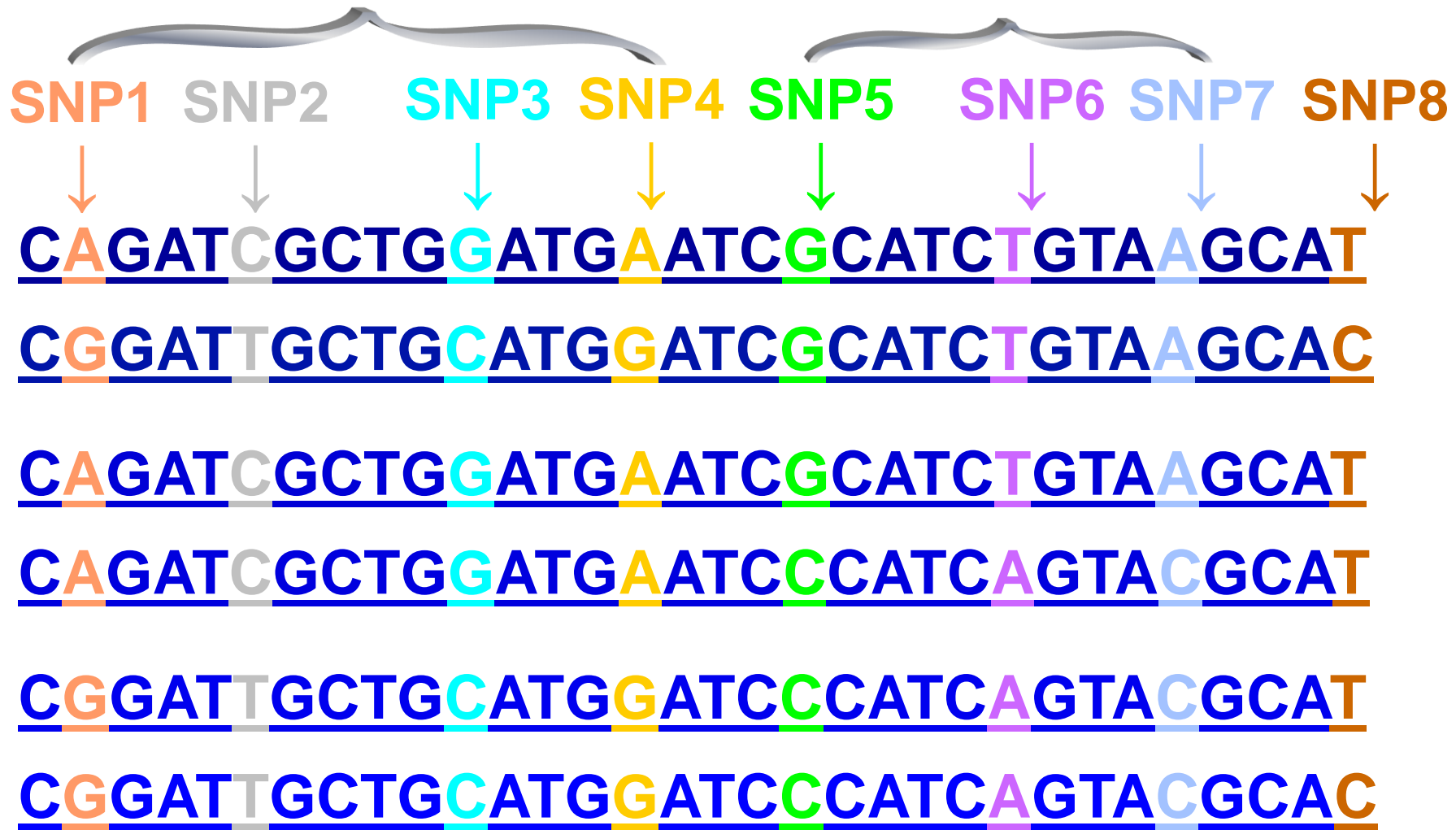
SNP 2 can predict SNP 3

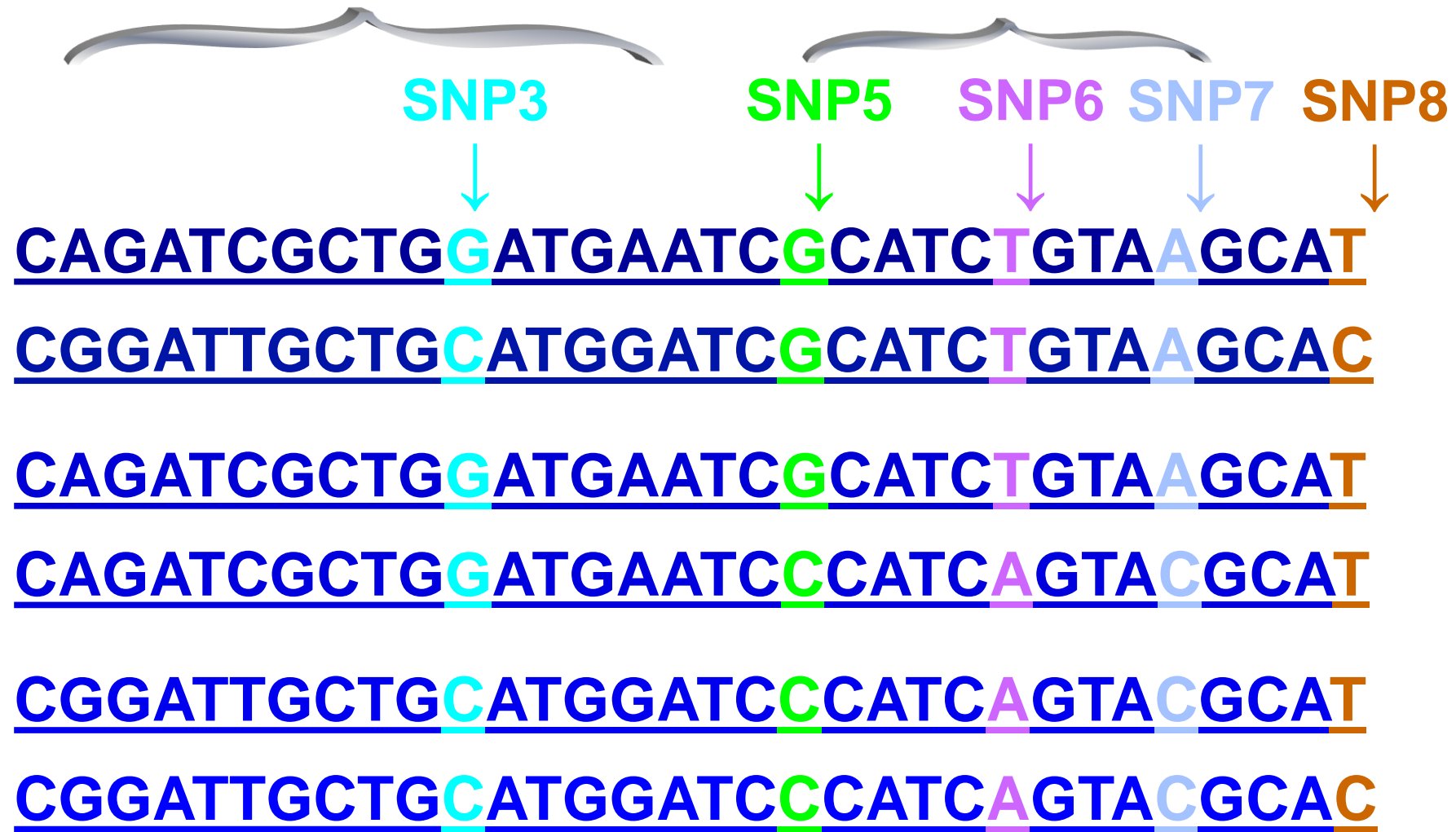
SNP 3 can predict SNP 2

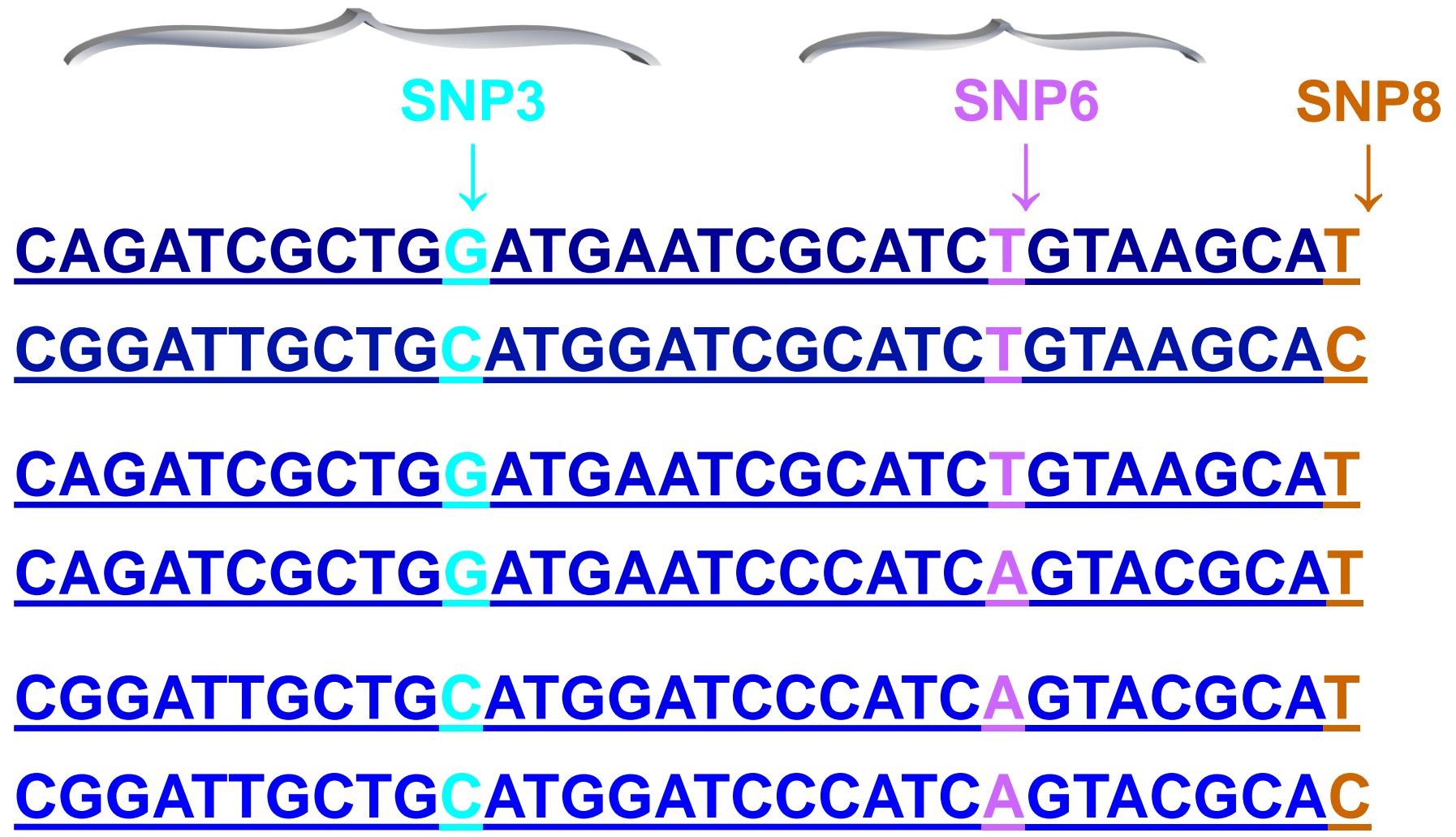
SNP 3 can predict SNP 4

Hap1	G	T	A	G
Hap2	C	T	A	T
Hap3	G	G	T	T
SNP #	1	2	3	4

SNPs 1 and 3 together predict SNP 4







GTT	35%
CTC	30%
GTT	10%
GAT	8%
CAT	7%
CAC	6%

other haplotypes **4%**

Three SNPs predict 96% different haplotypes

The Tagging problem

- **Given** a sample S of genotypes from a population P ; each sample has m SNPs
- **Find** positions of k ($k < m$) tag SNPs
- **Such that** one can reconstruct genotype g on all m SNPs in P from its restriction g' on k tag SNPs with certain accuracy

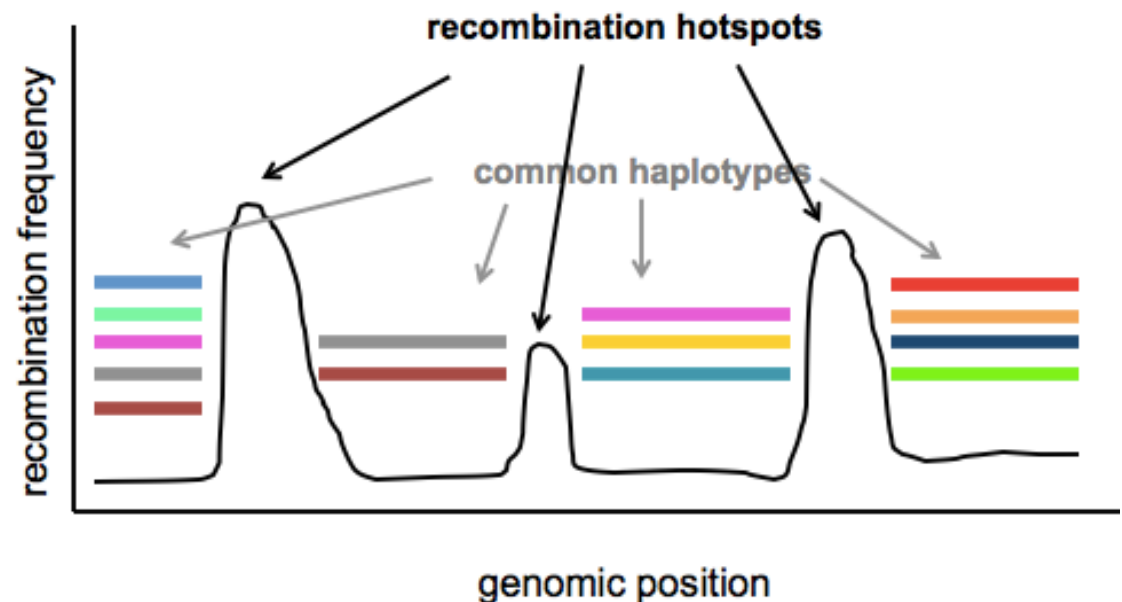
General framework of a tagging method

(Halldorsson et al., 2004)

1. Define a genomic region to search for tag SNPs.
2. Define a quality metric that quantifies how well a set of tag SNPs capture all the variance in the full data set.
3. Design an algorithm that selects a minimal number of tag SNPs that meet a desired quality threshold or optimizes the quality metric (as an objective function).

Define a search region

- Haplotype-block-based vs block-free methods
- Human genome consists of haplotype blocks (Daly et al., 2001; Dawson et al., 2002; Gabriel et al., 2002; Patil et al., 2001; Wall & Pritchard 2003).



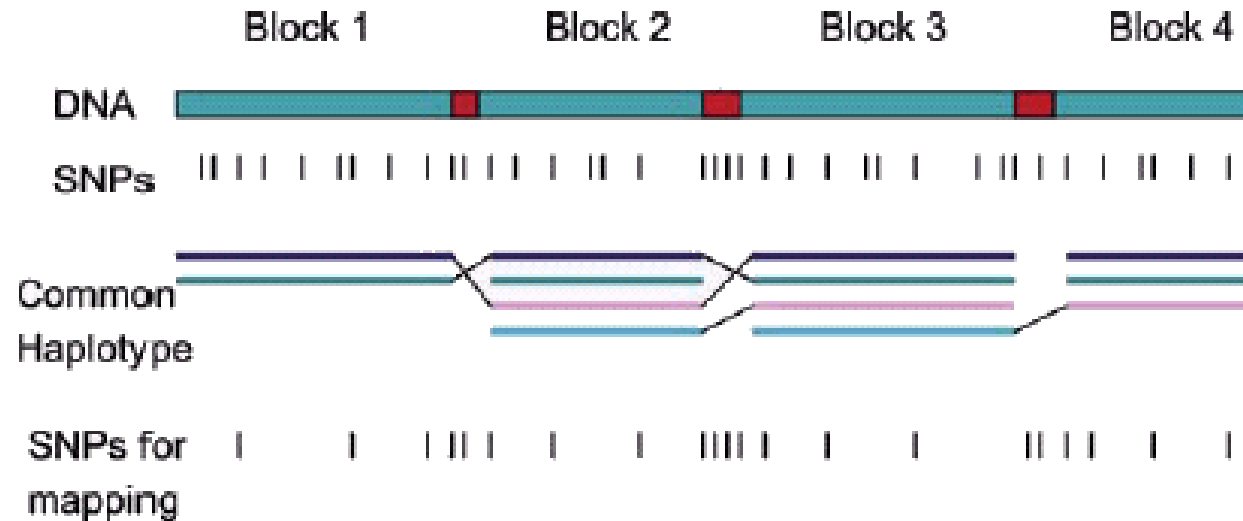
Block-based tagging

- Find a small set of SNPs **in each block** that captures the majority of SNP variation and identity common haplotypes **in that block**.
- But what exactly is a haplotype block?
 - High LD inside
 - Low haplotype diversity
 - Little recombination
 - ...

No consensus on a practical definition

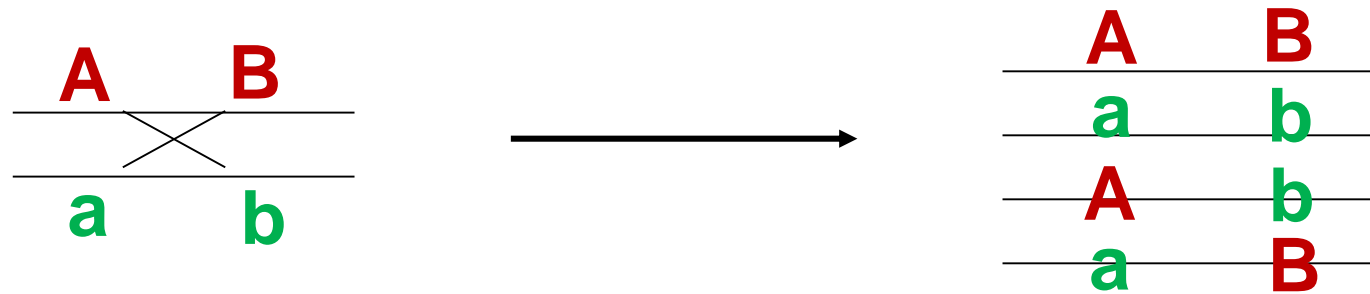
Block: Low haplotype diversity

- Patil et al., 2001
 - In each block, at least a certain proportion of observed or inferred haplotypes should be common haplotypes.



Block: no historical recombination

- Wang et al. 2002
 - A set of consecutive SNPs form a block if there is no historical recombination events (based on the four-gamete test)



< 4 haplotypes, $D' = 1$ \longrightarrow block

4 haplotypes, $D' < 1$ \longrightarrow boundary

Block: strong pairwise LD inside

- Gabriel et al. 2002
 - Blocks are partitioned based on whether the upper and lower confidence bounds on pairwise D' meet certain thresholds.
 - Specifically, the proportion of SNP pairs with strong LD (upper confidence bound $> .98$ and lower bound $> .7$) must account for at least 95% of all SNP pairs

Problems

- Block boundaries are ambiguous: they are sensitive to block definition and marker density (Boundary SNPs are often SNPs within recombination hotspots; until today they are not well tagged. Fine mapping is often needed.)
- Haplotype blocks are assumed to be independent, but adjacent blocks can still have substantial correlation.
- Not all genomic regions fit the haplotype-block model (Wall and Pritchard 2003).

Block-free Tagging

- Search for tag SNPs in a predefined neighborhood of each target SNP
- It is non-trivial to define the neighborhood (a sliding window).
 - There is usually an upper bound on the distance between a tag SNP and a target SNP (i.e., the maximal size of the window)
 - A small fixed window size (Meng et al., 2003)
 - A dynamically adjusted window size based on local LD extent (Halldorsson et al., 2004)

Define a quality metric

- Pairwise vs multivariate metrics
- LD measures (e.g., D' , r^2)
 - Select tags until a r^2 threshold (often > 0.8) is exceeded for every pair of target and tag SNPs (Carlson et al., 2004; Zhang and Jin, 2003)
 - Select the “best N ” tags by the number of target SNPs they can surrogate at a given r^2 (de Bakker et al., 2005)
 - The power to directly detect a causal SNP in Nr^2 samples is equivalent to the power to detect it indirectly (via markers) in N samples (Pritchard & Przeworski 2001).

Define a quality metric (cont.)

- Haplotype R^2 (Stram et al., 2003; Weale et al., 2003)
 - Extension of r^2 to Haplotypes
 - R_h^2 stands for the correlation between the frequency of haplotype h inferred from tag SNPs and all SNPs
- Statistical power (Genin 2001; Hu et al., 2004)
 - Assume, one at a time, that every SNP could be the disease mutation, which is unknown, and calculate pairwise power between the putative causal SNP and other SNPs
- Classic multivariate statistics used in PCA (Meng et al., 2003; Lin & Altman 2004), clustering (Ao 2005), or regression (He 2006)

Define a quality metric (cont.)

- Haplotype diversity
 - Coverage of common haplotypes (Patil et al, 2001; Zhang et al., 2002)
 - Coverage of overall haplotype diversity (Johnson et al., 2001)
 - “Informativeness” (Halldorsson et al., 2004)
 - Entropy (Hampe et al., 2003; Zhang et al., 2005)
 - If there are n haplotypes and the frequency of haplotype i is denoted by p_i , then the entropy of these haplotypes is defined as $S = -\sum_{i=1}^n p_i \log p_i$

Problems

- Not all the metrics have clear implications on the power-efficiency trade-off of association studies.
- Using pairwise metrics tend to overestimate the required number of tag SNPs
- Using multivariate metrics must deal with the fact that haplotypes are often unknown and need to be inferred.
- These metrics are based on one SNP or one block. The values need to be appropriately combined for genome-wide SNP selection.

Design an optimizing algorithm

- Computing the optimal solution to selecting the most informative SNPs is generally NP-hard (Bafna et al, 2003).
- Existing tagging methods use greedy (Carlson et al., 2004) or branch-and-bound (Avi-Itzhak et al., 2003).
- Dynamic programming is also applied (Zhang et al., 2002, 2003, 2004; Halldorsson et al., 2004).

Comparison of tagging methods

- Pairwise vs multivariate metrics
 - Multi-marker tagging tends to have fewer tags but more missed signals
- There is a lack of consistency across SNP sets selected by different methods, whether or not LD was present (Ding & Kullo, 2007; Goode et al., 2007).
- Quality metrics may not be as important to performance as optimizing algorithms.

Problems

- SNPs that are rare or have low r^2 with others are poorly tagged.
- Tagging loses its cost-saving advantage in regions of low LD.
- Tagging can be inaccurate when there is population stratification and allele frequencies are significantly different in subpopulations.
- Controversy exists over the extent to which tag SNPs (and GWAS) can help explore untyped structural polymorphism.
- Are these problems caused by tagging methods' dependency on LD? What other information can we use to find out the correlation of SNPs? What about genealogy? Can we find a set of tag SNPs such that a coalescent model can be as well simulated by these SNPs alone as by all SNPs?

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Thank you